Endothelial function and cardiovascular disease: Effects of quercetin and wine polyphenols

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Abstract

Endothelial dysfunction is an early pathophysiological feature and independent predictor of poor prognosis in most forms of cardiovascular diseases. Epidemiological studies report an inverse association between dietary flavonoid consumption and mortality from cardiovascular diseases. In the present paper, we review the effects of flavonoids, especially quercetin and wine polyphenols, on endothelial function and dysfunction and its potential protective role in hypertension, ischemic heart disease and stroke. *In vitro* studies show that flavonoids may exert multiple actions on the NO-guanylyl cyclase pathway, endothelium-derived hyperpolarizing factor(s) and endothelin-1 and protect endothelial cells against apoptosis. *In vivo*, flavonoids prevent endothelial dysfunction and reduce blood pressure, oxidative stress and end-organ damage in hypertensive animals. Moreover, some clinical studies have shown that flavonoid-rich foods can improve endothelial function in patients with hypertension and ischemic heart disease. Altogether, the available evidence indicates that quercetin and wine polyphenols might be of therapeutic benefit in cardiovascular diseases even though prospective controlled clinical studies are still lacking.

Keywords: Flavonoid, endothelial dysfunction, cardiovascular disease, hyperpolarizing factors

Introduction

Flavonoids represent the major class of polyphenolics [1]. The term flavonoid comprises several thousand plant-derived compounds sharing a common skeleton of phenylchromane. This basic structure allows a multitude of substitution patterns leading to several flavonoid subclasses such as flavonols, flavones, flavanones, catechins, anthocyanidins, isoflavones, dihydroflavonols and chalcones. They are widely distributed in the plant kingdom, being present in variable amounts in dietary fruits, vegetables, nuts, seeds, herbs, spices, tea and red wine [2]. Human daily intake of flavonols plus flavones has been estimated to be 22–23 mg. However, there are large variations within the population depending on their nutritional habits [3]. The interest in dietary flavonoids has grown in the last fifteen years after the publication of several epidemiological studies showing an inverse correlation between dietary consumption of flavonols and flavones and reduced incidence and mortality from cardiovascular disease and cancer [4,5]. For instance, the meta-analysis of seven prospective cohort studies concluded that the individuals in the top third of dietary flavonol intake are

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associated with a reduced risk of mortality from coronary heart disease as compared with those in the bottom third, after adjustment for known risk factors and other dietary components [6]. A limited number of intervention studies with flavonoids and flavonoid containing foods and extracts has been performed in several pathological conditions [7].

The first biological property described for flavonoids was related to the barrier function of the endothelium [8]. Due to their effect on capillary permeability, flavonoids were formerly considered vitamins. The term "vitamin P" (for Permeability) [8] was discontinued in the 1950s [9]. In recent years, a large amount of experimental and some clinical data have accumulated regarding the effects of flavonoids on the endothelium under physiological and pathological conditions. In the present paper we review the effects of flavonoids on endothelial function and its potential protective role in cardiovascular diseases. It must be pointed out that flavonoids represent a large group of different compounds and their effects may differ both quantitatively and qualitatively. Most of the available data regard flavonoid-rich foods (cocoa) or beverages (wine, juices and tea), extracts from wine or fruit juices and some isolated compounds such as the flavonols quercetin and kaemfperol, cathechins and the antocyanin delphinidin. The effects of isoflavone phytoestrogens derived from soy, genistein and daidzein, have been also widely studied, but differ from the most classic flavonoids in multiple ways and will not be analyzed in the present paper. The present review mainly focuses on the vascular effects of quercetin (representing a 60-75% of the total dietary flavonols plus flavones intake [4]) and wine polyphenolics (a complex mixture of different compounds), which have been widely studied by several research groups including ours and probably represent the most prototypical examples of a flavonoid and a flavonoid-rich extract, respectively.

Endothelial function and dysfunction

Endothelial factors and its physiological role

Endothelial cells, strategically located in the interface between the blood and vascular smooth muscle cells, release a number of vasoactive substances and play a major role in the control of vascular structure and function and platelet aggregation [10]. Under physiological conditions, the predominant effect of substances released by the endothelium is vasodilator, antiproliferative and antiaggregant, limiting the raise in blood pressure, regulating tissue blood flow and maintaining blood fluidity. Endothelial-derived substances with vasodilator and antiproliferative effects include NO, prostacyclin and endothelium-derived hyperpolarizing factors (EDHF), and substances with vasoconstrictor and mitogenic effects include endothelin-1 (ET-1) and PGH_{2 α}.

Endothelial dysfunction

The equilibrium between vasodilator and vasoconstrictors is shifted in cardiovascular diseases, where vasoconstrictor and proliferative effects predominate, leading to hypertension, atherosclerosis, platelet aggregation and ischemia. Thus, endothelial dysfunction is characterized by impaired endotheliumdependent vasodilatation and a prothrombotic and proinflammatory state of endothelial cells. Endothelial dysfunction is an early and independent predictor of poor prognosis in most forms of cardiovascular diseases [11-13]. Thus, alterations in endothelial function have been consistently found in hypertension, atherosclerosis, coronary heart disease, diabetes, sepsis, obesity and aging [11-14].

Nitric oxide

NO is formed from L-arginine by the endothelial NO synthase (eNOS) which requires $Ca^{2+}/calmodulin$, FAD, FMN and tetrahydrobiopterin (BH4) as cofactors [15]. eNOS is activated upon increases in cytosolic Ca^{2+} which occurs in response to multiple stimuli such as shear stress, hormones, platelet derived substances and several drugs. NO diffuses to the adjacent smooth muscle cells where it activates soluble guanylyl cyclase and induces vasodilatation [16]. It also exerts antiaggregant effects in platelets. NO plays a fundamental role in controlling blood pressure, tissue flow and blood fluidity.

The most characteristic pathophysiological feature of endothelial dysfunction is a diminished bioactivity of endothelium-derived NO resulting in impaired vascular homeostasis [11-14]. The classical approach to analyze endothelial function both in vitro and in vivo is to stimulate endothelial NO release with agonists such as acetylcholine. The mechanisms involved in the decreased endothelial-derived NO responses have been extensively studied and include [11,17,18] : (1) inhibition of the signal transduction from receptor activation to eNOS activation, (2) changes in the activity and/or expression of eNOS, (3) changes in the vascular levels of superoxide anion (O_2^-) and, thus, O_2^- -driven NO inactivation (see below) and, (4) changes in the sensitivity to the NO-cyclic guanosine monophosphate (cGMP) pathway in vascular smooth muscle cells. Reduced NO synthesis associated to endothelial dysfunction may be caused by reduced expression of eNOS, postransductional modification of the enzyme (e.g. phosphorylation or fatty acid modifications), interactions with heat shock protein 90 (hsp90), calmodulin or caveolin, suboptimal concentrations of the substrate L-arginine or the cofactor tetrahydrobiopterin (BH4), or the presence of endogenous NOS inhibitors such as asymmetric dimethylarginine and N-monometylarginine [11]. The isoprenoid geranylgeranyl pyrophosphate, an intermediate factor in the cholesterol synthesis pathway, also inhibits the activity of eNOS. In addition, hyperhomocysteinemia, which is associated with increased risk of stroke, ischemic heart disease, peripheral vascular disease and venous thrombosis, leads to reduced eNOS activity [19].

Oxidative stress

Oxidative stress, characterized by an increased endogenous production of reactive oxygen species such as O_2^- and hydrogen peroxide (H₂O₂), is a central cause of endothelial dysfunction. Potential sources of vascular O₂⁻ production include membrane NADPHdependent oxidase, xanthine oxidase, cyclooxygenase, lipoxygenase, the mitochondrial respiratory chain and eNOS [18]. NADPH oxidase is a multi-subunit enzymatic complex [20] which comprises membranesubunits $(p22^{phox} and gp91^{phox} [nox 2] or its [nox 1 and$ nox 4]) and cytoplasmic subunits (p47^{phox}, p67^{phox} and p40^{phox}). This complex is considered to be the most important source of O₂⁻ in the vessel wall [21-23]. Interestingly, excess of O_2^- generation synthesized by NADPH oxidase is critically involved in the breakdown of NO associated to endothelial dysfunction in experimental and clinical hypertension [11,18,24]. Under certain circumstances, eNOS becomes dysfunctional ("uncoupled") and electrons are diverted to molecular oxygen rather than to Larginine, producing O₂⁻ instead of NO. eNOS uncoupling represents another potential mechanism involved in endothelial dysfunction and it may result from a deficiency of L-arginine, BH₄ or may be promoted by the presence of its endogenous inhibitors. Thus, eNOS uncoupling is also a critical mechanism in experimental and clinical endothelial dysfunction [25].

 O_2^- can interfere with NO by a number of mechanisms, both with its synthesis and its activity, ultimately leading to reduced endothelium-dependent vasodilatation [18]. Acutely, O_2^- reacts with and inactivates very rapidly NO, to form peroxynitrite (ONOO⁻) [26]. O_2^- also oxidizes BH₄, a cofactor required for NO synthesis. Beside, O_2^- and its by-products, ONOO⁻ and H₂O₂, are involved in the oxidation of LDL cholesterol and in the inflammatory process accompanying atherosclerosis. Oxidized LDL, in turn, increases the synthesis of caveolin-1, which inactivates eNOS. Moreover, O_2^- and H₂O₂ may produce direct vasoconstrictor effects [27].

Cyclooxygenase

Activation of endothelial cyclooxygenase leads to the transformation of arachidonic acid into PGH_2 that is metabolized by several enzymes into different products [28]. In endothelial cells, under physiological conditions, the major metabolite is PGI_2 [29]. The vasodilator activity of PGI_2 is determined by

the expression of specific receptors on vascular smooth muscle cells that are coupled to adenylyl cyclase [30]. Cyclooxygenases exist in two isoforms, COX-1 and COX-2, which are constitutively expressed or inducible, respectively [31,32]. Some of the cyclooxygenase by-products are endothelium-derived contracting factors. Indeed, the use of inhibitors of cyclooxygenase, like indomethacin, pointed out an increased participation of endothelial cyclooxygenase-derived vasoconstrictor metabolites in conductance and resistance arteries in animal models of cardiovascular disease and in humans [33-35]. The cyclooxygenase metabolite(s) involved in the increased endotheliumdependent vasoconstriction are those who can activate the thromboxane A₂(TXA₂)/endoperoxide receptor such as PGH₂, TXA₂ or PGF_{2 α} [33–37]. Hence, the use of an antagonist of the TXA2/endoperoxide receptor improves endothelial dysfunction in certain circumstances [33-37]. The increased participation of vasoconstrictor factors derived from cyclooxygenase has been associated with an increased expression of COX-1 and COX-2 proteins in the vessel wall [34]. Regarding the cyclooxygenase isoform responsible for the release of vasoconstrictor prostanoids, no consensus can yet be established. The change in the pattern of products released by cyclooxygenase might be related to an alteration in the lipid substrates. Indeed, an increase in lipid peroxidation was associated with an increased participation of cyclooxygenase-derived vasoconstrictors in endotheliumdependent relaxation in the rat. This may be a consequence of an increased oxidative stress on the cyclooxygenase-dependent response for example in the course of aging. Finally, it should be noted that NO can chemically interact with PGH₂ [38,39] and the observed increased production of cyclooxygenasederived EDCF may account for the decreased availability of NO.

EDHF

Another important endothelium-derived relaxing factor, especially in resistance arteries, is EDHF. The nature of EDHF, depending on the type of artery considered, has been proposed to be epoxyeicosatrienoic acid (EET), K^+ , anandamide and H_2O_2 [40–42]. The responses mediated by EDHF are resistant to NO synthase and cyclooxygenase inhibitors but are sensitive to the combination of the SK_{Ca} and IK_{Ca} channel inhibitors apamin plus charibdotoxin. EDHFmediated responses are initiated by the activation of endothelial SK_{Ca} and IK_{Ca} channels which leads to the hyperpolarization of endothelial cells [42]. The consecutive hyperpolarization of smooth muscle cells involves the spread of an electric current through myoendothelial gap junctions [42]. Endothelial dysfunction is associated in some cases with a reduced EDHF-component of the relaxation, independently

or in addition to NO. Possible reasons to explain the differences observed in the magnitude of the alteration in EDHF-mediated relaxation could be species or anatomical heterogeneity of vasomotor regulation conductance vs. resistance arteries. Also changes might occur downstream of EDHF release such as a decreased expression of voltage- and Ca²⁺-activated K⁺ channels [43]. Under physiological conditions, these channels serve as an hyperpolarizing force that oppose contraction. Thus, their reduced expression could lead to a decreased vasodilatory capacity. However, this change is not uniform among different vascular beds and species.

ET-1

An increased participation of the potent vasoconstrictor ET-1 could also explain reduction of endotheliumdependent vasodilatation. Indeed, endothelial dysfunction is associated with an upregulation of mRNA expression of the precursor of ET-1, preproendothelin-1 and ET-1 protein [44,45]. Numerous conditions characterized by an impaired availability of NO have been found to be associated with enhanced synthesis of ET-1 and vice-versa, thereby suggesting that these two factors have a reciprocal regulation [46]. ET-1 was described to exert a bidirectional effect by either enhancing NO production via ET_B receptors located in endothelial cells or blunting its effect via ET_A receptors prevalently located in the vascular smooth muscle cells. Conversely, NO was found to inhibit ET-1 synthesis in different cell types [46]. Several factors affect in opposite direction the transcription of prepro-ET-1 and NOS genes, peroxisome proliferator-activated receptors playing a key role in these regulatory mechanisms [46]. All these data suggest that endothelial dysfunction is also characterized by this dual effect on the NO and ET-1 pathways.

In vitro effects of flavonoids

Vascular effects of several flavonoids have been widely studied in vitro. However, caution should be made and there are some limitations for the relevance of results from in vitro studies because little is known on bioavailability of flavonoids. Quercetin and related flavonoids are absorbed in appreciable amounts in the small intestine. However, they are metabolized both at the enterocytes and in the liver into methylated and glucurono- and sulfo-conjugated derivatives which show a long-lasting presence in plasma [2]. Unconjugated levels of flavonoids in plasma are low and it is currently believed that most of the in vivo effects of flavonoids may be due to the their conjugated metabolites. Unfortunately, the literature on the vascular effects of these metabolites is scarce, and it is presently unknown which of the effects obtained *in vitro* reported for the parent compounds are also pertinent to the metabolites.

Direct effect on vascular smooth muscle tone and proliferation

Quercetin and related flavonoids exert direct (endothelium-independent) vasodilator effects in isolated arteries [47–49]. This vasodilator effect occurs similarly in arteries constricted by different stimuli including endogenous vasoconstrictors (noradrenaline, ET-1 or TXA₂), PKC activators (phorbol esters) and depolarizing agents (KCl). The molecular mechanisms of the direct vasodilator effects are not definitely established but might involve multiple actions on protein kinases such as MLCK and PKC. Interestingly, quercetin and its metabolites are more potent in coronary arteries [50] and in resistance than in conductance vessels [51].

Several studies have shown that quercetin and red wine polyphenols inhibit proliferation and hypertrophy of vascular smooth muscle cells in culture [52-54]. The inhibitory effects of quercetin on DNA synthesis of vascular smooth muscle cells stimulated by TNF- α appear to be related to reduced ERK1/2 activity [52], a kinase playing a major role in cell proliferation and differentiation. In contrast, in angiotensin II-stimulated vascular smooth muscle cells, quercetin and its conjugated glucuronide inhibit hypertrophy via down-regulation of the JNK pathway [53]. Quercetin blocks cell-cycle in G1-phase, and subsequently down-regulates cyclins and CDKs and up-regulates the CDK inhibitor p21 expression in vascular smooth muscle cells [53].

Effects on nitric oxide

Flavonoids exert multiple effects on endothelialderived NO. In addition to its direct vasodilator effects, grape juices and wine or extracts from a variety of vegetables and fruits have been reported to induce endothelium-dependent vasorelaxation [49,55-58]. Endothelium-dependent relaxation has also been reported for several isolated flavonoids, especially the anthocyanin delphinidin [56] and the flavone chrysin [59]. In contrast, for other structurally related anthocyanins (e.g. malvidin and cyanidin) and other flavonols (e.g. quercetin and kaempferol), a possible endothelium-dependent relaxation is masked by endothelium-independent effects [47-49,51,56]. Endothelium-dependent relaxation induced by chrisin, red wine polyphenols and delphinidin is inhibited by NO synthase inhibitors, indicating that it was mediated by the NO-cyclic GMP pathway. Furthermore, this effect does not appear to be related to a protective effect on NO but rather to an increase of NO synthesis. These flavonoids increased endothelial cytosolic Ca²⁺ levels, [55-58] the physiological

signal for eNOS activation. The increase in Ca^{2+} in endothelial cells was inhibited by a mixture of superoxide dismutase and catalase suggesting the involvement of O_2^- [60].

In addition, by scavenging O_2^- or by inhibiting its synthesis (see below), flavonoids protect NO from O₂⁻-driven inactivation, increasing its half-life and its biological activity [61]. Due to its antioxidant properties, flavonoids can potentially avoid BH4 oxidation and eNOS uncoupling. Therefore, under conditions of high O_2^- , flavonoids potentiate NO- and endothelium-dependent relaxation, reverting oxidative stress-induced endothelial dysfunction. However, the chemical relationships between flavonoids and NO is more complex because flavonoids may also scavenge NO [62]. This reaction involved the autooxidation of flavonoids in aqueous buffers producing O_2^- which ultimately inactivates NO [61]. However, the rate of NO scavenging of flavonoids including quercetin is much lower than that of the physiological NO scavenger haemoglobin.

NO exerts its vasodilator effects by activating soluble guanylyl cyclase in vascular smooth muscle cells and the subsequent increase in cGMP [16]. In turn, cGMP is metabolized by cyclic nucleotide phosphodiesterases (PDEs) and thus, NO activity and endotheliumdependent relaxation are strongly dependent on PDE activity. Therefore, PDE inhibitors can prevent endothelial dysfunction in some circumstances [63]. Several flavonoids have also been reported to inhibit several PDE isoforms [64]. In fact, some flavonoids (e.g. kaempferol) potentiate the relaxant response to the guanylyl cyclase activator sodium nitroprusside [47]. Thus, inhibition of PDEs may represent another potential mechanism for flavonoid-induced prevention of endothelial dysfunction.

Flavonoids may also regulate NO activity at the level of eNOS mRNA and/or protein expression. Longterm incubation of endothelial cells with red wine or the anthocyanins delphinidin, malvidin, cyanidin and paeonidin increased eNOS expression while most isolated flavonols were without effect [65–66].

Antioxidant effects

The health-promoting effects of flavonoids are usually attributed to their powerful antioxidant activity. Quercetin and related flavonoids, in the low micromolar range, are effective scavengers of several reactive oxygen species including O_2^- , singlet oxygen and hydroxyl and peroxyl radicals [67–68]. Furthermore, the glucuronized metabolites of quercetin also behave as antioxidants [69]. The copper chelating properties of flavonoids have also been suggested to contribute to its antioxidant effect [70]. Besides these ROS scavenging effects, and probably more importantly, flavonoids inhibit several ROS generating enzymes including xanthine oxidase [71] and the membrane NADPH

oxidase complex in neutrophils [72]. As mentioned above, by reducing cellular O_2^- concentrations flavonoids protect NO and increase its bioactivity. In addition, they are powerful inhibitors of LDL oxidation, a key event in the process of atherosclerotic plaque genesis. The hydrophilic properties of flavonoids facilitate its localization at the water phase-lipid bilayer interface and thereby protect lipids and vitamin E in phospholipid bilayers and in LDL from the initial attack by aqueous radicals. In addition, flavonoids prevent cytotoxicity of oxidized LDL.

On the other hand, flavonoids are not devoid of prooxidant properties. For example, quercetin can autooxidize in aqueous solutions to generate free radicals [61] and may also deplete intracellular thiols such as glutathione both *in vitro* and *in vivo*. The pro-oxidant effect of red wine polyphenolics may also play a positive role in endothelial function because $O_2^$ generation in endothelial cells can stimulate Ca²⁺ signalling and enhance eNOS activity (see above) and increase EDHF release (see below).

Effects on EDHF

An additional mechanism by which flavonoids might induce endothelium-dependent relaxation is by stimulating the release of EDHF. In fact, red wine polyphenols have been shown to produce endothelium-dependent relaxation and hyperpolarization in porcine coronary artery rings in the presence of NOS and cyclooxygenase inhibitors [73]. These responses were inhibited by the combination of charybdotoxin plus apamin and also reduced by antioxidants, membrane permeant analogues of superoxide dismutase, and diphenylene iodonium, an inhibitor of flavin-dependent enzymes. These findings demonstrate that red wine polyphenols cause EDHFmediated relaxations of coronary arteries involving a pro-oxidant mechanism. The flavone chrysin has also been suggested to release EDHF in addition to NO in the perfused mesenteric bed [74].

Effects on ET-1

The effect of flavonoids on the ET-1 system has been studied by Corder and colleagues [75]. A low concentrations of red wine polyphenols is sufficient to strongly inhibit ET-1 release and transcription of prepro-endothelin-1 in bovine aortic endothelial cells (BAEC). None of the purified compounds (quercetin, resveratrol, catechin, epicathechin, pelargonidin, delphinidin, cyanidin) possess such property at this concentration. The mechanism appears to involve tyrosine kinases. In contrast, other study reports that quercetin ($0.5-50 \mu$ M) is able to inhibit ET-1 release in human endothelial umbilical vein [76]. No data are available for the *in vivo* effect of flavonoids on ET-1 production.

Effects on endothelial cell apoptosis and proliferation

Apoptosis of endothelial cells strongly affect endothelium permeability and thus facilitates the development of various pathologies. With regard to the endothelial cells, polyphenols exert a double-edged role in apoptosis by acting on multiple pathways. Polyphenols have been shown to exert protective effects in vitro against apoptosis mediated by oxidized LDL and hydrogen peroxide in BAEC and fibroblasts [77]. The inhibition by polyphenols of the apoptosis induced by oxidized LDL in BAEC is related to the modulation of the calcium homeostasis. Recently, we demonstrate that delphinidin is able to protect endothelial cells against apoptosis [78]. Of particular interest is the finding that the antiapoptotic effect of delphinidin results from increased eNOS expression via mitogenactivated protein kinase inhibitor-sensitive pathway. The effect of delphinidin also involves the NO and guanylyl cyclase-dependent pathway and is associated with the maintenance of endothelial cytosolic calcium level in a physiological range and the decrease of cytochrome c release from the mitochondria. Flavonoids affect apoptosis partly through caspase pathway. Indeed, polyphenols from oolong tea, theasinensin A, promote apoptosis of tumor cells by activating caspases 3 and 9 [79]. Besides, polyphenols can modulate the level of expression of protective proteins (Bcl-2, Bxl-xL, A1) or proapoptotic factors (Bax, Bid, Bak) [80]. Whatever the effects of flavonoids on endothelial, they can either be pro- or anti-apoptotic agents in order to maintain endothelial integrity depending on the aggression.

Few studies were conducted on the effect of flavonoids on endothelial proliferation but in general they are reported as inhibitors of this process [81]. We recently reported that endothelial cell migration and proliferation are prevented by delphinidin through the involvement of cyclin D1- and A-dependent pathway [78,82]. Surprisingly, the effect of delphinidin does not involve the NO pathway in contrast to its action on endothelial cell apoptosis.

Ex vivo and *in vivo* effects of flavonoids: Experimental and clinical

Endothelial function in hypertension

Blood pressure elevation is a common and powerful contributor to all of the major cardiovascular diseases, including coronary disease, stroke, peripheral artery disease, renal disease and heart failure [83]. The blood pressure lowering effects of vegetables and fruit-rich diet in hypertensive patients is well known [84]. It has been mainly attributed to the presence of multiple antioxidants present in these foodstuffs. The effects of the flavonol quercetin and red wine polyphenols have been analyzed in detail in several animal models of hypertension.

Chronic quercetin treatment (5 or 10 mg Kg^{-1} day⁻¹) lowered blood pressure or prevented the development of hypertension in all experimental models tested including spontaneously hypertensive rats (SHR), nitric oxide deficient rats (chronically treated with L-NAME), desoxycorticosterone acetate-(DOCA) salt hypertensive rats, Dahl salt-sensitive hypertensive rats and two-kidney, one-clip Goldblatt hypertensive rats [85-89]. Therefore, regardless of the status of the renin-angiotensin system, volumeexpansion, nitric oxide, renal injury or oxidative stress, quercetin produced consistent antihypertensive effects. Quercetin also exerted end-organ protection in hypertensive animals, reducing the cardiac and renal hypertrophy, the proteinuria and the vascular remodelling associated with elevated blood pressure. These preventive effects on structural changes may result from the reduction in blood pressure and the subsequent reduced trophic effect of haemodynamic forces but also by a direct antiproliferative effect of quercetin as discussed above. These experimental models, as well as human essential hypertension, are associated with reduced endothelium derived NO-dependent vasodilatation. In SHR, DOCA-salt and Goldblatt rats, chronic quercetin restored the impaired endothelial vasodilator function as measured by the relaxant response to acetylcholine. Increased urinary NOx (nitrites + nitrates, main NO metabolites) was also found. All these models were associated with increased plasma, vascular and hepatic oxidative status as measured by plasma, tissue and urinary levels of either malonyldialdehyde or isoprostanes, and quercetin consistently reduced these parameters [85-87,89]. Altogether these results suggest a role for reduced O_2^- -driven NO inactivation. Furthermore, in a recent study, quercetin has been shown to downregulate the expression of SHR aortic p47^{phox}, a key regulatory subunit of NADPH oxidase which was accompanied by a reduction in vascular NADPH oxidase derived O_2^- production [90]. Additionally, apparent changes in endothelial-dependent vasodilatation may occur as a consequence of the opposing effects of the release of endothelium-derived vasoconstrictor prostanoids [91,92]. Chronic quercetin did not modify the endothelium-dependent vasoconstriction in SHR [85,90] but markedly inhibited the later in NO-deficient [86] and in Goldblatt rats [89]. In addition, aortic TXA₂ production was increased in SHR [89], in NO-deficient [86] and in Goldblatt rats [89] as compared to their normotensive controls. Again, quercetin reduced aortic TXA₂ levels in NO-deficient and in Goldblatt rats but not in SHR. Other isolated flavonoids such as baicalein and flavone have been reported to prevent the development of endothelial dysfunction in SHR [93]. Moreover, baicalein

reduced blood pressure in hypertensive animals via an increase in prostacyclin production [94].

It must be noted that quercetin had minimal, if any, effects on blood pressure or endothelial function in normotensive animals. This is consistent with the lack of effect on blood pressure of a high dose of quercetin in normotensive subjects [95] or the minimal effect of a fruit and vegetables rich diet in the subgroup of normotensive subjects [84]. Thus, quercetin and related flavonoids may exert their effect on endothelial function specifically when endothelial function is impaired and blood pressure is elevated.

Administration of ProvinolsTM (a standardized red wine polyphenolics extract) reduced the increase in both blood pressure and protein synthesis in heart and aorta caused by chronic inhibition of NO synthesis [96]. This was accompanied by reduced end-organ damage such as myocardial fibrosis and aortic thickening. ProvinolsTM also prevented endothelium-dysfunction that was associated with an increase of NOS activity, a moderate enhancement of endothelial NOS expression and a reduction of oxidative stress in the left ventricle and aorta. In addition, an alcohol-free grape skin extract also significantly reduced systolic, mean and diastolic arterial pressure in DOCA-salt and L-NAME-induced experimental hypertension in rats [97].

To the best of our knowledge, there are no studies published in hypertensive patients with isolated flavonoids. However, there are some reports on the effects of flavonoid-rich foods such as cocoa and fruit juices compared to similar foods or beverages devoid or poor in flavonoids. The effects of flavonoid-rich dark chocolate compared to white chocolate on blood pressure and endothelial function in essential hypertension have been analyzed in a double-blind cross-over study [98]. Dark chocolate significantly reduced blood pressure and insulin resistance and improved flow-dilated (endothelium-dependent) vasodilatation. Another doubleblind, cross-over study conducted in 12 hypertensive patients which received alternately high-flavonoid sweetie juice and low-flavonoid sweetie juice for 5 weeks [99] showed that the flavonoid rich juice significantly reduced blood pressure. Even when concerns can be raised about which specific components are responsible for these effects, these well controlled trials suggest that flavonoid-rich foods exert a beneficial effect on blood pressure and endothelial function in hypertensive patients. Interestingly, in healthy male adults the ingestion of flavanol-rich cocoa was associated with increased levels of circulating NO species and enhanced endothelium-dependent vasodilation and these effects were mimicked by (-)-epicatechin [100].

LDL oxidation and atherosclerosis

As indicated above, one of the mechanisms responsible for endothelial dysfunction is the increased LDL oxidation and flavonoids possess the ability to reduce lipid sensitivity to oxidation. In animals, Vinson et al. [101] reported that polyphenols from red wine or grape juice reduce the plasmatic concentration of lipids in hamsters. This is consistent with studies in humans. Red, but not white, wine consumption for 2 weeks resulted in a 20% reduction in the propensity of plasma to undergo lipid peroxidation in healthy subjects [102]. In addition, short-term ingestion of purple grape juice has been shown to reduce LDL susceptibility to oxidation in patients with coronary artery disease [103]. This is also in harmony with the recent study in hypercholesterolemic postmenopausal women with red wine complementation [104]. However, in several studies, the LDL oxidation was not modified by flavonoid intake in human. de Rijke et al. [105] did not observe a modification of the LDL oxidizability in healthy volunteers after red wine consumption (550 ml during 4 weeks). O'Reilly et al. [106] have published that the intake of flavonoids from onions and green tea does not modify the level of LDL oxidation in human. Moreover, a study in postmenopausal women have recently determined that the intake of lyophilized grape powder (rich in flavans, anthocyanins, quercetin, myricetin, kaempferol, and resveratrol) during 4 weeks does not reduce the LDL oxidation, even when plasma triglycerides, plasma LDL-cholesterol and apolipoproteins B and E concentrations, are lowered by the treatment [107]. Therefore, the effect of flavonoids on LDL oxidation may vary depending on their structure, the type of natural diet they were originated and the dose used. In addition, several flavonoids have been reported to prevent the development of atherosclerosis lesions in ApoE deficient mice without effects on plasma LDL or HDL cholesterol levels [108].

Coronary disease

The pathophysiology of ischemic heart disease and acute coronary syndromes involves multiple alterations in vascular reactivity, vascular structure, and interactions of the vessel wall with circulating blood elements [109]. Alterations in the serum lipid profile and the development of atherosclerosis in the coronary arteries is an early and crucial event in ischemic heart disease. Furthermore, blood pressure elevation is a powerful risk factor for coronary disease. Controlled trials indicate that a reduction of LDL cholesterol and atherosclerotic lesions and blood pressure produces a dramatic decline in the risk of coronary disease [83,109,110]. Therefore, the antiatherogenic and antihypertensive effects of flavonoids (as discussed above) might prevent coronary disease. Most acute coronary events result from a rupture in the atherosclerotic plaque and the subsequent thrombus formation. Degradation of the interstitial collagen conferring biomechanical strength to the plaque fibrous cap by matrix metalloproteinases (MMPs) appears to be involved in the plaque instability and its rupture. Coronary vasospasm may also contribute to acute impaired arterial flow. Flavonoids exert platelet antiaggregant properties [111,112], reduce the expression of matrix metalloproteinases (MMP-2 and MMP-9) [113] and induce a coronary vasodilator effect [50,114] providing additional potential mechanisms for the prevention of the acute coronary events.

Endothelial dysfunction in coronary artery disease was reported as early as 1986 [115]. Reduced bioavailability of endothelial NO is involved in the genesis progression and complications of coronary atherosclerosis. An increase in endothelium-dependent vasodilatation has been observed in patients with coronary disease treated with grape purple juice or tea [103,116]. Post-ischemic reperfusion occurring in coronary diseases is generally associated with a reduction of endogenous NO production resulting from endothelial dysfunction and tissue damage linked to neutrophil infiltration. The capacity of flavonoids to enhance NO generation probably plays a crucial role to prevent ischemia. These observations have been reinforced by the abrogation of the protective effect of wine polyphenols after blockade of NO production with NO synthase inhibitor. Experimental studies in animal models have shown that grape juice, red wine or isolated polyphenols including flavone and quercetin reduce the contractile dysfunction of the heart and protect against the cellular lesions induced by cardiac ischemia [117-120]. These effects may be observed following the oral intake of these substances or after their perfusion in an isolated heart before the induction of an ischemia. Using an ex vivo rat model of cardiac ischemia/ reperfusion, we have recently published that shortterm red wine polyphenol extract treatment protects against post-ischemic infarction via decreased oxidative stress and implies an involvement of NOdependent pathway [121].

Stroke

The primary cause of stroke is an interruption of cerebral blood flow, which occurs either during an arterial or venous obstruction (embolus or local thrombosis) or during a cardiac arrest. The pathophysiological processes in stroke are complex and depend on the severity, duration and localization of the ischemic damage in the brain. Flavonoids can be potentially used in both preventive and acute approaches to the treatment of strokes [122]. Indeed, flavonoids prevent endothelial dysfunction responsible for atherosclerosis and thrombosis and lower blood pressure and blood cholesterol. With regard to grape and red wine, the main reported studies examining their *in vivo* effects on cerebral ischemia have been conducted using resveratrol [122].

The beneficial effect of resveratrol on cerebral ischemia is discussed in a recent review and is out of the scoop of the present review which deals mainly on wine and grape extracts [122]. Dietary supplementation of grape powder or an extract of the powder improved ischemia-induced delayed neuron death and glial cell activation. The grape extract showed effective protection even when it was given after ischemia [123] Quercetin has been reported to be able to scavenge superoxide anions released during reperfusion after forebrain ischemia using a four-vessel occlusion model in rats [124]. Consequently, quercetin (30 mg/kg intraperitoneally) given after middle cerebral artery ligation in a permanent middle cerebral artery occlusion model in rats significantly decreased the infarct volume. We have investigated the potential protective effects of wine polyphenols in a rat model of transient cerebral ischemia [125,126]. A red wine polyphenolic extract was given orally during one week before the induction of the stroke. Microdialysis analysis shows that polyphenols do not modify the energy metabolism and the oxidative stress, but are able to reduce the excitotoxicity by inhibiting the massive release of glutamate and aspartate. The cerebral blood flow has also been monitored and polyphenols can improve the residual blood flow both during occlusion and early reperfusion. Furthermore, polyphenols may induce vascular remodeling, with an increase of the internal diameter of the brain vessels. Most interestingly, wine polyphenols reduce brain infarct size an effect that is associated with inhibition of ischemia-induced excitotoxicity and improvement of cerebral blood flow. Data from this study provide an experimental basis supporting the view that wine polyphenols may be beneficial for stroke protection.

Summary and conclusions

The present review mainly focuses on the vascular effects of quercetin and wine polyphenols. A typical commercial bottle of red wine contains approximately 1.8 g/L of total polyphenols. So the total amount of polyphenols found in a glass of red wine is about 200 mg. Flavonoids constitute the majority of these phenols in red wine and their concentrations are ranged from 1365 to 1500 mg/L [127].

Endothelial dysfunction is a common pathophysiological feature which develops early in the evolution of cardiovascular diseases. The severity of endothelial dysfunction is related to poor prognosis and interventions reducing cardiovascular risk have been shown to improve endothelial function. A large body of evidence supports the hypothesis that flavonoids show protective effects *in vitro* on the vessel wall, particularly on endothelial function, involving multiple mechanisms. In endothelial cells, they induce the release of vasoactive factors such as NO and EDHF and prevent the apoptosis induced by oxidative damage. They also inhibit the release or the action of endothelial derived constrictor factors such as ET-1. In vascular smooth muscle cells, flavonoids directly inhibit the contractile and proliferative response induced by several pathological stimuli. In several animal models of endothelial dysfunction associated to hypertension, flavonoids improve endothelial function, reduce blood pressure and the associated cardiac, vascular and renal damage and reduce the expression and the activity of prooxidative enzymes such as membrane NADPH oxidase. A limited number of clinical studies have also shown that flavonoid rich foods can improve endothelial function in patients with hypertension, atherosclerosis and coronary heart disease.

Outstandingly, flavonoids can potentially interact with most of the steps involved in the pathophysiology of ischemic heart disease and stroke. Flavonoids might prevent ischemic heart disease by multiple mechanisms operating simultaneously in both the chronic (e.g. by inhibiting the genesis and development of atherogenic plaques, hypertension and endothelial dysfunction) and the acute phase (e.g. by inhibiting coronary vasoconstriction, plaque vulnerability, platelet adhesion and aggregation and myocardial necrosis). Similarly, flavonoids act on different phases of stroke. For the acute phase, flavonoids improve cerebral blood flow, prevent platelet aggregation and thrombosis, reduce excitotoxicity and inhibit oxidative stress. For the intermediate phase, flavonoids reduce inflammation and protect endothelial integrity. For the late phase, flavonoids interfere with ischemia induced cell death mechanisms such as apoptosis and necrosis.

Altogether, the available evidence indicates that flavonoids might be of therapeutic benefit in cardiovascular diseases, which are still a major public health problem. Nevertheless, prospective controlled clinical studies with flavonoids are still lacking. These human intervention studies would be necessary before establishing any recommendations about dietary habits or administration of dietary supplements.

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References

- Rice-Evans CA, Packer L. Flavonoids in health and disease. New York: Marcel Dekker Inc. 1998.
- [2] Manach C, Scalbert A, Morand C, Remesy C, Jimenez L. Polyphenols: Food sources and bioavailability. Am J Clin Nutr 2004;79:727–747.
- [3] Hertog MG, Kromhout D, Aravanis C, Blackburn H, Buzina R, Fidanza F, Giampaoli S, Jansen A, Menotti A, Nedeljkovic S. Flavonoid intake and long-term risk of coronary heart disease

and cancer in the seven countries study. Arch Intern Med 1995;155:381-386.

- [4] Hertog MG, Feskens EJ, Hollman PC, Katan MB, Kromhout D. Dietary antioxidant flavonoids and risk of coronary heart disease: The Zutphen elderly study. Lancet 1993;342:1007–1011.
- [5] Birt DF, Hendrich S, Wang W. Dietary agents in cancer prevention: Flavonoids and isoflavonoids. Pharmacol Ther 2001;90:157-177.
- [6] Huxley RR, Neil HA. The relation between dietary flavonol intake and coronary heart disease mortality: A meta-analysis of prospective cohort studies. Eur J Clin Nutr 2003;57:904–908.
- [7] Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. Am J Clin Nutr 2005;811(Suppl):2438–255S.
- [8] Rusznyak S, Szent-Györgyi A. Vitamin P: Flavonols as vitamins. Nature 1936;138:27–29.
- [9] Joint Committee on Nomenclature of the American Society of biological chemists -American Institute of nutrition. Term "vitamin P" recommended to be discontinued. Science 1950;112:268.
- [10] Vane JR, Anggard EE, Botting RM. Regulatory functions of the vascular endothelium. N Engl J Med 1990;323:27–36.
- [11] Endemann DH, Schiffrin EL. Endothelial dysfunction. J Am Soc Nephrol 2004;15:1983–1992.
- [12] Schachinger V, Britten MB, Zeiher AM. Prognostic impact of coronary vasodilator dysfunction on adverse long-term outcome of coronary heart disease. Circulation 2000;101: 1899–1906.
- [13] Widlansky ME, Gokce N, Keaney JF, Vita JA. The clinical implications of endothelial dysfunction. J Am Coll Cardiol 2003;42:1149–1160.
- [14] Kawashima S, Yokoyama M. Dysfunction of endothelial nitric oxide synthase and atherosclerosis. Arterioscler Thromb Vasc Biol 2004;24:998–1005.
- [15] Moncada S, Palmer RM, Higgs EA. Nitric oxide: Physiology, pathophysiology and pharmacology. Pharmacol Rev 1991;43:109–142.
- [16] Warner TD, Mitchell JA, Sheng H, Murad F. Effects of cyclic GMP on smooth muscle relaxation. Adv Pharmacol 1994;26:171–194.
- [17] Miwa Y, Hirata K, Kawashima S, Akita H, Yokoyama M. Lysophosphatidylcholine inhibits receptor-mediated Ca²⁺ mobilization in intact endothelial cells of rabbit aorta. Arterioscler Thromb Vasc Biol 1997;17:1561–1567.
- [18] Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: The role of oxidant stress. Circ Res 2000;87:840-844.
- [19] Lentz A, Rodionov RN, Dayal S. Hyperhomocysteinemia, endothelial dysfunction, and cardiovascular risk: The potential role of ADMA. Atheroscler Suppl 2003;4:61–65.
- [20] Babior BM. NADPH oxidase: An update. Blood 1999;93:1464–1476.
- [21] Perez-Vizcaino F, Lopez-Lopez JG, Santiago R, Cogolludo A, Zaragoza-Arnaez F, Moreno L, Alonso MJ, Salaices M, Tamargo J. Postnatal maturation in nitric oxide-induced pulmonary artery relaxation involving cyclooxygenase-1 activity. Am J Physiol Lung Cell Mol Physiol 2002;283:L839–L848.
- [22] Griendling KK, Sorescu D, Ushio-Fukai M. NADP;H oxidase: Role in cardiovascular biology and disease. Circ Res 2000;86:494–501.
- [23] Calo LA, Davis PA, Pagnin E, Semplicini A, Pessina AC. NADPH oxidase, superoxide overproduction and nitric oxide bioavailability in essential hypertension. J Hypertens 2005;23:665–666.

- [24] Virdis A, Neves MF, Amiri F, Touyz RM, Schiffrin EL. Role of NADPH oxidase on vascular alterations in angiotensin IIinfused mice. J Hypertens 2004;22:535–542.
- [25] Vasquez-Vivar J, Kalyanaraman B, Martasek P. The role of tetrahydrobiopterin in superoxide generation from eNOS: Enzymology and physiological implications. Free Radic Res 2003;37:121–127.
- [26] Koppenol WH. The basic chemistry of nitrogen monoxide and peroxynitrite. Free Rad Biol Med 1998;25:385–391.
- [27] Jin L, Ying Z, Webb RC. Activation of Rho/Rho kinase signaling pathway by reactive oxygen species in rat aorta. Am J Physiol Heart Circ Physiol 2004;287:H1495-H1500.
- [28] Terlain B, Jouzeau JY, Gillet P, Lecompte T, Netter P. Inducible cyclooxygenase. New relationships between nonsteroidal anti- inflammatory agents and inhibition of synthesis of prostaglandins. Presse Med 1995;24:491–496.
- [29] Moncada S, Vane JR. Pharmacology and endogenous roles of prostaglandin endoperoxides, thromboxane A2, and prostacyclin. Pharmacol Rev 1978;30:293–331.
- [30] Narumiya S, Sugimoto Y, Ushikubi F. Prostanoid receptors: Structures, properties, and functions. Physiol Rev 1999;79:1193–1226.
- [31] Mitchell JA, Warner TD. Cyclo-oxygenase-2: Pharmacology, physiology, biochemistry and relevance to NSAID therapy. Br J Pharmacol 1999;128:1121–1132.
- [32] Fitzgerald GA. Cardiovascular pharmacology of nonselective nonsteroidal anti- inflammatory drugs and coxibs: Clinical considerations. Am J Cardiol 2002;89:26D-32D.
- [33] Koga T, Takata Y, Kobayashi K, Takishita S, Yamashita Y, Fujishima M. Ageing suppresses endothelium-dependent relaxation and generates contraction mediated by the muscarinic receptors in vascular smooth muscle of normotensive Wistar-Kyoto and spontaneously hypertensive rats. J Hypertens Suppl 1988;6:S243–S245.
- [34] Matz RL, de Sotomayor MA, Schott C, Stoclet JC, Andriantsitohaina R. Vascular bed heterogeneity in agerelated endothelial dysfunction with respect to NO and eicosanoids. Br J Pharmacol 2000;131:303–311.
- [35] Taddei S, Virdis A, Mattei P, Ghiadoni L, Fasolo CB, Sudano I, Salvetti A. Hypertension causes premature aging of endothelial function in humans. Hypertension 1997;29:736–743.
- [36] Kung CF, Luscher TF. Different mechanisms of endothelial dysfunction with aging and hypertension in rat aorta. Hypertension 1995;25:194–200.
- [37] Heymes C, Habib A, Yang D, Mathieu E, Marotte F, Samuel J, Boulanger CM. Cyclo-oxygenase-1 and -2 contribution to endothelial dysfunction in ageing. Br J Pharmacol 2000;131:804–810.
- [38] Ito T, Kato T, Iwama Y, Muramatsu M, Shimizu K, Asano H, Okumura K, Hashimoto H, Satake T. Prostaglandin H2 as an endothelium-derived contracting factor and its interaction with endothelium-derived nitric oxide. J Hypertens 1991;9:729–736.
- [39] Auch-Schwelk W, Katusic ZS, Vanhoutte PM. Nitric oxide inactivates endothelium-derived contracting factor in the rat aorta. Hypertension 1992;195:442–445.
- [40] Campbell WB, Harder DR. Endothelium-derived hyperpolarizing factors and vascular cytochrome P450 metabolites of arachidonic acid in the regulation of tone. Circ Res 1999;84:484–488.
- [41] Dora KA, Garland CJ. Properties of smooth muscle hyperpolarization and relaxation to K⁺ in the rat isolated mesenteric artery. Am J Physiol Heart Circ Physiol 2001;280:H2424-H2429.
- [42] Busse R, Edwards G, Feletou M, Fleming I, Vanhoutte PM, Weston AH. EDHF: Bringing the concepts together. Trends Pharmacol Sci 2002;23:374–380.
- [43] Marijic J, Li Q, Song M, Nishimaru K, Stefani E, Toro L. Decreased expression of voltage- and Ca²⁺-activated K⁺

channels in coronary smooth muscle during aging. Circ Res 2001;88:210–216.

- [44] Barton M. Endothelial dysfunction and atherosclerosis: Endothelin receptor antagonists as novel therapeutics. Curr Hypertens Rep 2000;2:84–91.
- [45] Brunner F, Bras-Silva C, Cerdeira AS, Leite-Moreira AF. Cardiovascular endothelins: Essential regulators of cardiovascular homeostasis. Pharmacol Ther 2006; in press.
- [46] Rossi GP, Seccia TM, Nussdorfer GG. Reciprocal regulation of endothelin-1 and nitric oxide: Relevance in the physiology and pathology of the cardiovascular system. Int Rev Cytol 2001;209:241–272.
- [47] Duarte J, Perez-Vizcaino F, Utrilla P, Jimenez J, Tamargo J, Zarzuelo A. Vasodilatory effects of flavonoids in rat aortic smooth muscle. Structure-activity relationships. Gen Pharmacol 1993;24:857–862.
- [48] Duarte J, Perez-Vizcaino F, Zarzuelo A, Jimenez J, Tamargo J. Vasodilator effects of quercetin on isolated rat vascular smooth muscle. Eur J Pharmacol 1993;239:1–7.
- [49] Fitzpatrick DF, Hirschfield SL, Coffey RG. Endotheliumdependent vasorelaxing activity of wine and other grape products. Am J Physiol 1993;265:H774–H778.
- [50] Ibarra M, Perez-Vizcaino F, Cogolludo A, Duarte J, Zaragoza-Arnaez F, Lopez-Lopez JG, Tamargo J. Cardiovascular effects of isorhamnetin and quercetin in isolated rat and porcine vascular smooth muscle and isolated rat atria. Planta Med 2002;68:307–310.
- [51] Perez-Vizcaino F, Ibarra M, Cogolludo AL, Duarte J, Zaragoza-Arnaez F, Moreno L, Lopez-Lopez JG, Tamargo J. Endothelium-independent vasodilator effects of the flavonoid quercetin and its methylated metabolites in rat conductance and resistance arteries. J Pharmacol Exp Ther 2002; 302:66–72.
- [52] Moon SK, Cho GO, Jung SY, Gal SW, Kwon TK, Lee YC, Madamanchi NR, Kim CH. Quercetin exerts multiple inhibitory effects on vascular smooth muscle cells: Role of ERK1/2, cell-cycle regulation and matrix metalloproteinase-9. Biochem Biophys Res Commun 2003;301:1069–1078.
- [53] Yoshizumi M, Tsuchiya K, Suzaki Y, Kirima K, Kyaw M, Moon JH, Terao J, Tamaki T. Quercetin glucuronide prevents VSMC hypertrophy by angiotensin II via the inhibition of JNK and AP-1 signaling pathway. Biochem Biophys Res Commun 2002;293:1458–1465.
- [54] Iijima K, Yoshizumi M, Hashimoto M, Kim S, Eto M, Ako J, Liang YQ, Sudoh N, Hosoda K, Nakahara K, Toba K, Ouchi Y. Red wine polyphenols inhibit proliferation of vascular smooth muscle cells and downregulate expression of cyclin A gene. Circulation 2000;101:805–811.
- [55] Andriambeloson E, Kleschyov AL, Muller B, Beretz A, Stoclet JC, Andriantsitohaina R. Nitric oxide production and endothelium-dependent vasorelaxation induced by wine polyphenols in rat aorta. Br J Pharmacol 1997; 120:1053–1058.
- [56] Andriambeloson E, Magnier C, Haan-Archipoff G, Lobstein A, Anton R, Beretz A, Stoclet JC, Andriantsitohaina R. Natural dietary polyphenolic compounds cause endothelium-dependent vasorelaxation in rat thoracic aorta. J Nutr 1998; 128:2324–2333.
- [57] Stoclet JC, Kleschyov A, Andriambeloson E, Diebolt M, Andriantsitohaina R. Endothelial no release caused by red wine polyphenols. J Physiol Pharmacol 1999;50:535–540.
- [58] Andriambeloson E, Stoclet JC, Andriantsitohaina R. Mechanism of endothelial nitric oxide-dependent vasorelaxation induced by wine polyphenols in rat thoracic aorta. J Cardiovasc Pharmacol 1999;33:248–254.
- [59] Duarte J, Jimenez R, Villar IC, Perez-Vizcaino F, Jimenez J, Tamargo J. Vasorelaxant effects of the bioflavonoid chrysin in isolated rat aorta. Planta Med 2001;67:567–569.

- [60] Duarte J, Andriambeloson E, Diebolt M, Andriantsitohaina R. Wine polyphenols stimulate superoxide anion production to promote calcium signaling and endothelial-dependent vasodilatation. Physiol Res 2004;53:595–602.
- [61] Lopez-Lopez G, Moreno L, Cogolludo A, Galisteo M, Ibarra M, Duarte J, Lodi F, Tamargo J, Perez-Vizcaino F. Nitric oxide NO; scavenging and NO protecting effects of quercetin and their biological significance in vascular smooth muscle. Mol Pharmacol 2004;65:851–859.
- [62] Van Acker SA, Tromp MN, Haenen GR, Van der Vijgh WJ, Bast A. Flavonoids as scavengers of nitric oxide radical. Biochem Biophys Res Commun 1995;214:755-759.
- [63] Vlachopoulos C, Tsekoura D, Alexopoulos N, Panagiotakos D, Aznaouridis K, Stefanadis C. Type 5 phosphodiesterase inhibition by sildenafil abrogates acute smoking-induced endothelial dysfunction. Am J Hypertens 2003;17:1040–1014.
- [64] Picq M, Dubois M, Prigent AF, Nemoz G, Pacheco H. Inhibition of the different cyclic nucleotide phosphodiesterase isoforms separated from rat brain by flavonoid compounds. Biochem Int 1989;18:47–57.
- [65] Wallerath T, Poleo D, Li H, Forstermann U. Red wine increases the expression of human endothelial nitric oxide synthase: A mechanism that may contribute to its beneficial cardiovascular effects. J Am Coll Cardiol 2003;41:471–478.
- [66] Wallerath T, Li H, Godtel-Ambrust U, Schwarz PM, Forstermann U. A blend of polyphenolic compounds explains the stimulatory effect of red wine on human endothelial NO synthase. Nitric Oxide 2005;12:97–104.
- [67] O'Reilly JD, Sanders TA, Wiseman H. Flavonoids protect against oxidative damage to LDL *in vitro*: Use in selection of a flavonoid rich diet and relevance to LDL oxidation resistance *ex vivo*? Free Radic Res 2000;33:419–426.
- [68] Ozgova S, Hermanek J, Gut I. Different antioxidant effects of polyphenols on lipid peroxidation and hydroxyl radicals in the NADPH-: Fe-ascorbate- and Fe-microsomal systems. Biochem Pharmacol 2003;66:1127–1237.
- [69] Moon JH, Tsushida T, Nakahara K, Terao J. Identification of quercetin 3-O-beta-D-glucuronide as an antioxidative metabolite in rat plasma after oral administration of quercetin. Free Radic Biol Med 2001;30:1274–1285.
- [70] Roland A, Patterson RA, Leake DS. Measurement of copperbinding sites on low density lipoprotein. Arterioscler Thromb Vasc Biol 2001;21:594–602.
- [71] Chang WS, Lee YJ, Lu FJ, Chiang HC. Inhibitory effects of flavonoids on xanthine oxidase. Anticancer Res 1993;13: 2165–2170.
- [72] Tauber AI, Fay JR, Marletta MA. Flavonoid inhibition of the human neutrophil NADPH-oxidase. Biochem Pharmacol 1984;33:1367–1369.
- [73] Ndiaye M, Chataigneau T, Andriantsitohaina R, Stoclet JC, Schini-Kerth VB. Red wine polyphenols cause endothelium dependent EDHF-mediated relaxations in porcine coronary arteries via a redox-sensitive mechanism. Biochem Biophys Res Commun 2003;310:371–377.
- [74] Villar IC, Galisteo M, Vera R, O'Valle F, Garcia-Saura MF, Zarzuelo A, Duarte J. Effects of the dietary flavonoid chrysin in isolated rat mesenteric vascular bed. J Vasc Res 2004;41:509–516.
- [75] Corder R, Douthwaite JA, Lees DM, Khan NQ, Viseu Dos Santos AC, Wood EG, Carrier MJ. Endothelin-1 synthesis reduced by red wine. Nature 2001;414:863–864.
- [76] Zhao X, Gu Z, Attele AS, Yuan CS. Effects of quercetin on the release of endothelin, prostacyclin and tissue plasminogen activator from human endothelial cells in culture. J Ethnopharmacol 1999;67:279–285.
- [77] Vieira O, Escargueil-Blanc I, Meilhac O, Basile JP, Laranjinha J, Almeida L, Salvayre R, Negre-Salvayre A. Effect of dietary phenolic compounds on apoptosis of human

cultured endothelial cells induced by oxidized LDL. Br J Pharmacol 1998;123:565–573.

- [78] Martin S, Giannone G, Andriantsitohaina R, Martinez MC. Delphinidin, an active compound of red wine, inhibits endothelial cell apoptosis via nitric oxide pathway and regulation of calcium homeostasis. Br J Pharmacol 2003;139:1095–1102.
- [79] Pan MH, Liang YC, Lin-Shiau SY, Zhu NQ, Ho CT, Lin JK. Induction of apoptosis by the oolong tea polyphenol theasinensin A through cytochrome c release and activation of caspase-9 and caspase-3 in human U937 cells. J Agric Food Chem 2000;48:6337–6346.
- [80] Nam S, Smith DM, Dou QP. Tannic acid potently inhibits tumor cell proteasome activity, increases p27 and Bax expression, and induces G1 arrest and apoptosis. Cancer Epidemiol Biomarkers Prev 2001;10:1083–1088.
- [81] Igura K, Ohta T, Kuroda Y, Kaji K. Resveratrol and quercitin inhibit angiogenesis *in vitro*. Cancer Lett 2001;171:11–16.
- [82] Favot L, Martin S, Keravis T, Andriantsitohaina R, Lugnier C. Involvement of cyclin-dependent pathway in the inhibitory effect of delphinidin on angiogenesis. Cardiovasc Res 2003;59:479–487.
- [83] Kannel WB. Risk stratification in hypertension: New insights from the Framingham Study. Am J Hypertens 2000;13: 3S-10S.
- [84] Appel LJ, Moore TJ, Obarzanek E, Vollmer WM, Svetkey LP, Sacks FM, Bray GA, Vogt TM, Cutler JA, Windhauser MM, Lin PH, Karanja N. A clinical trial of the effects of dietary patterns on blood pressure. N Engl J Med 1997;33: 1117–1124.
- [85] Duarte J, Perez-Palencia R, Vargas F, Ocete MA, Perez-Vizcaino F, Zarzuelo A, Tamargo J. Antihypertensive effects of the flavonoid quercetin in spontaneously hypertensive rats. Br J Pharmacol 2001;133:117–124.
- [86] Duarte J, Jimenez R, O'Valle F, Galisteo M, Perez-Palencia R, Vargas F, Perez-Vizcaino F, Zarzuelo A, Tamargo J. Protective effects of the flavonoid quercetin in chronic nitric oxide deficient rats. J Hypertens 2002;20:1843–1854.
- [87] Galisteo M, Garcia-Saura MF, Jimenez R, Villar IC, Wangensteen R, Zarzuelo A, Vargas F, Duarte J. Effects of quercetin treatment on vascular function in deoxycorticosterone acetate-salt hypertensive rats. Comparative study with verapamil. Planta Med 2004;70:334–341.
- [88] Aoi W, Niisato N, Miyazaki H, Marunaka Y. Flavonoidinduced reduction of ENaC expression in the kidney of Dahl salt-sensitive hypertensive rat. Biochem Biophys Res Commun 2004;315:892–896.
- [89] García-Saura MF, Galisteo M, Villar IC, Bermejo A, Zarzuelo A, Vargas F, Duarte J. Effects of chronic quercetin treatment in experimental renovascular hypertension. Mol Cell Biochem 2005;270:147–155.
- [90] Sanchez M, Galisteo M, Vera R, Villar IC, Zarzuelo A, Tamargo J, Perez-Vizcaino F, Duarte J. Quercetin downregulates NADPH oxidase: Increases eNOS activity and prevents endothelial dysfunction in spontaneously hypertensive rats. J Hypertens 2006;24:75–84.
- [91] Auch-Schwelk W, Katusic ZS, Vanhoutte PM. Thromboxane A2 receptor antagonists inhibit endothelium-dependent contractions. Hypertension 1990;15:699–703.
- [92] Yang D, Feletou M, Levens N, Zhang JN, Vanhoutte PM. A diffusible substances;mediates endothelium-dependent contractions in the aorta of SHR. Hypertension 2003;41: 143–148.
- [93] Macha A, Mustafa MR. Chronic treatment with flavonoids prevents endothelial dysfunction in spontaneously hypertensive rat aorta. J Cardiovasc Pharmacol 2005;46:36–40.
- [94] Takizawa H, DelliPizzi AM, Nasjletti A. Prostaglandin I2 contributes to the vasodepressor effect of baicalein in hypertensive rats. Hypertension 1998;31:866–871.

- [95] Conquer JA, Maiani G, Azzini E, Raguzzini A, Holub BJ. Supplementation with quercetin markedly increases plasma quercetin concentration without effect on selected risk factors for heart disease in healthy subjects. J Nutr 1998;128: 593–597.
- [96] Pechanova O, Bernatova I, Babal P, Martinez MC, Kysela S, Stvrtina S, Andriantsitohaina R. Red wine polyphenols prevent cardiovascular alterations in L-NAME-induced hypertension. J Hypertens 2003;22:1551–1559.
- [97] Soares De Moura R, Costa Viana FS, Souza MA, Kovary K, Guedes DC, Oliveira EP, Rubenich LM, Carvalho LC, Oliveira RM, Tano T, Gusmao Correia ML. Antihypertensive, vasodilator and antioxidant effects of a vinifera grape skin extract. J Pharm Pharmacol 2002;54:1515–1520.
- [98] Grassi D, Necocione S, Lippi C, Croce G, Valeri L, Pasqualetti P, Desideri G, Blumberg JB, Ferri C. Cocoa reduces blood pressure and insulin resistance and improves endothelium-dependent vasodilation in hypertensives. Hypertension 2005;46:398-405.
- [99] Reshef N, Hayari Y, Goren C, Boaz M, Madar Z, Knobler H. Antihypertensive effect of sweetie fruit in patients with stage I hypertension. Am J Hypertens 2005;18:1360–1363.
- [100] Schroeter H, Heiss C, Balzer J, Kleinbongard P, Keen CL, Hollenberg NK, Sies H, Kwik-Uribe C, Schmitz HH, Kelm M. (-)-Epicatechin mediates beneficial effects of flavanol-rich cocoa on vascular function in humans. Proc Natl Acad Sci USA 2006;103:1024–1029.
- [101] Vinson JA, Teufel K, Wu N. Red wine, dealcoholized red wine, and especially grape juice, inhibit atherosclerosis in a hamster model. Atherosclerosis 2001;156:67–72.
- [102] Fuhrman B, Lavy A, Aviram M. Consumption of red wine with meals reduces the susceptibility of human plasma and low-density lipoprotein to lipid peroxidation. Am J Clin Nutr 1995;61:549–554.
- [103] Stein JH, Keevil JG, Wiebe DA, Aeschlimann S, Folts JD. Purple grape juice improves endothelial function and reduces the susceptibility of LDL cholesterol to oxidation in patients with coronary artery disease. Circulation 1999;100: 1050–1055.
- [104] Naissides M, Mamo JC, James AP, Pal S. The effect of chronic consumption of red wine on cardiovascular disease risk factors in postmenopausal women. Atherosclerosis 2005; in press.
- [105] de Rijke YB, Demacker PN, Assen NA, Sloots LM, Katan MB, Stalenhoef AF. Red wine consumption does not affect oxidizability of low-density lipoproteins in volunteers. Am J Clin Nutr 1996;63:329–334.
- [106] O'Reilly JD, Mallet AI, McAnlis GT, Young IS, Halliwell B, Sanders TA, Wiseman H. Consumption of flavonoids in onions and black tea: Lack of effect on F2-isoprostanes and autoantibodies to oxidized LDL in healthy humans. Am J Clin Nutr 2001;73:1040–1044.
- [107] Zern TL, Wood RJ, Greene C, West KL, Liu Y, Aggarwal D, Shachter NS, Fernandez ML. Grape polyphenols exert a cardioprotective effect in pre- and postmenopausal women by lowering plasma lipids and reducing oxidative stress. J Nutr 2005;135:1911–1917.
- [108] Hayek T, Fuhrman B, Vaya J, Rosenblat M, Belinky P, Coleman R, Elis A, Aviram M. Reduced progression of atherosclerosis in apolipoprotein E-deficient mice following consumption of red wine, or its polyphenols quercetin or catechin, is associated with reduced susceptibility of LDL to oxidation and aggregation. Arterioscler Thromb Vasc Biol 1997;17:2744–2752.
- [109] Libby P. Current concepts of the pathogenesis of the acute coronary syndromes. Circulation 2001;104:365–372.
- [110] Larosa JC, He J, Vupputuri S. Effect of statins on risk of coronary disease. JAMA 1999;282:2340–2346.

- [111] Gryglewski RJ, Korbut R, Robak J, Swies J. On the mechanism of antithrombotic action of flavonoids. Biochem Pharmacol 1987;36:317–322.
- [112] de Gaetano G, De Curtis A, di Castelnuovo A, Donati MB, Iacoviello L, Rotondo S. Antithrombotic effect of polyphenols in experimental models: A mechanism of reduced vascular risk by moderate wine consumption. Ann N Y Acad Sci 2004;957:174–188.
- [113] Huang YT, Hwang JJ, Lee PP, Ke FC, Huang JH, Huang CJ, Kandaswami C, Middleton E, Jr, Lee MT. Effects of luteolin and quercetin: Inhibitors of tyrosine kinase, on cell growth and metastasis-associated properties in A431 cells overexpressing epidermal growth factor receptor. Br J Pharmacol 1999;128:999–1010.
- [114] Rendig SV, Symons JD, Longhurst JC, Amsterdam EA. Effects of red wine, alcohol, and quercetin on coronary resistance and conductance arteries. J Cardiovasc Pharmacol 2001;38:219–227.
- [115] Ludmer PL, Selwyn AP, Shook TL, Wayne RR, Mudge GH, Alexander RW, Ganz P. Paradoxical vasoconstriction induced by acetylcholine in atherosclerotic coronary arteries. N Engl J Med 1986;315:1046–1051.
- [116] Duffy SJ, Keaney JF, Jr, Holbrook M, Gokce N, Swerdloff PL, Frei B, Vita JA. Short- and long-term black tea consumption reverses endothelial dysfunction in patients with coronary artery disease. Circulation 2001;104:151–156.
- [117] Cui J, Cordis GA, Tosaki A, Maulik N, Das DK. Reduction of myocardial ischemia reperfusion injury with regular consumption of grapes. Ann N Y Acad Sci 2002;957:302–307.
- [118] Sato M, Ray PS, Maulik G, Maulik N, Engelman RM, Bertelli AA, Bertelli A, Das DK. Myocardial protection with red wine extract. J Cardiovasc Pharmacol 2000;35:263–268.
- [119] Ning XH, Ding X, Childs KF, Bolling SF, Gallagher KP. Flavone improves functional recovery after ischemia in isolated reperfused rabbit hearts. J Thorac Cardiovasc Surg 1993;105:541–549.
- [120] Brookes PS, Digerness SB, Parks DA, Darley-Usmar V. Mitochondrial function in response to cardiac ischemiareperfusion after oral treatment with quercetin. Free Radic Biol Med 2002;32:1220–1228.
- [121] Ranaivo HR, Diebolt M, Andriantsitohaina R. Wine polyphenols induce hypotension, and decrease cardiac reactivity and infarct size in rats: Involvement of nitric oxide. Br J Pharmacol 2004;142:671-678.
- [122] Simonyi A, Wang Q, Miller RL, Yusof M, Shelat PB, Sun AY, Sun GY. Polyphenols in cerebral ischemia: Novel targets for neuroprotection. Mol Neurobiol 2005;31:135–148.
- [123] Wang Q, Simonyi A, Li W, Sisk BA, Miller RL, MacDonald RS, Lubahn DE, Sun GY, Sun AY. Dietary grape supplement ameliorates cerebral ischemia-induced neuronal death in gerbils. Mol Nutr Food Res 2005;49:443–451.
- [124] Dajas F, Rivera F, Blasina F, Arredondo F, Echeverry C, Lafon L, Morquio A, Heizen H. Cell culture protection and *in vivo* neuroprotective capacity of flavonoids. Neurotox Res 2003;5:425–432.
- [125] Curin Y, Ratajczak P, Dupont A, Ritz M-F, Mendelowitsch A, Pinet F, Andriantsitohaina R. Identification par analyse protéomique des déterminants moléculaires de la neuroprotection induite par les polyphenols dands l'ischémie cérébrale. Arch Mal Coeur Vaiss 2005;98:392.
- [126] Curin Y, Ritz M-F, Cam E, Andriantsitohaina R, Mendelowitsch A. Chronic treatment with red wine polyphenolic compounds protects against vascular and neuronal excitotoxicity in response to focal cerebral ischemia in male rats. Arch Mal Coeur Vaiss 2004;97:417.
- [127] Waterhouse AL. Wine phenolics. Ann NY Acad Sci 2002;957:21–36.